

Evaluation of Novel Anti-TNF α and IL-6R Therapies in a Rheumatoid Arthritis (RA) Quantitative Systems Pharmacology (QSP) Platform.

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Introduction

- RA is a common autoimmune disease associated with progressive disability, systemic complications, early death, and socioeconomic costs.
- RA is characterized by synovial inflammation and hyperplasia, autoantibody production, cartilage and bone destruction, and systemic features.
- Anti-TNFα, anti-IL-6, and anti-IL-6 receptor (IL-6R) therapies are approved for use in RA.
- A significant number of patients do not respond to treatment. Therefore, the **development** of new treatment approaches is warranted.

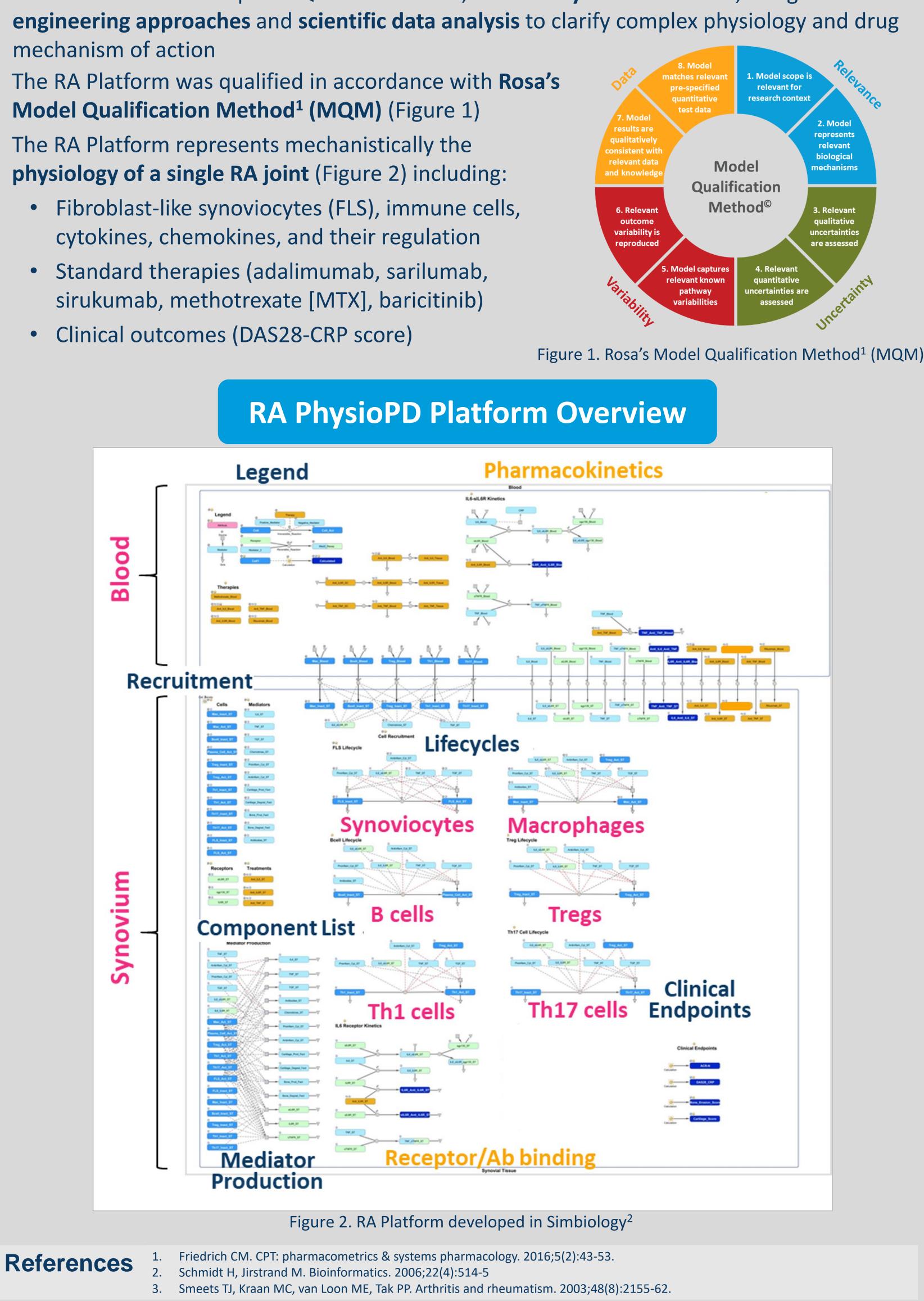
Objectives

- Support the development of **novel anti-TNFα and IL-6R therapies for RA**
- Gain a deeper understanding of the key biological pathways impacting clinical response to anti-TNF α or anti-IL-6R
- Identify RA patient phenotypes with optimal response to therapy

Methods

PhysioPD[™] Research Platforms are mechanistic, QSP models that allow investigation of the connection between mechanisms and outcomes.

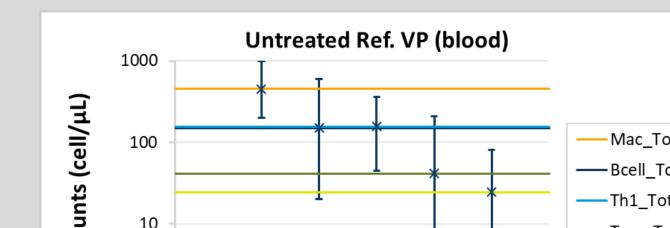
- Rosa and Sanofi developed a QSP model of RA, the RA PhysioPD Platform, using mechanism of action
- The RA Platform was qualified in accordance with **Rosa's** Model Qualification Method¹ (MQM) (Figure 1)
- The RA Platform represents mechanistically the
 - Fibroblast-like synoviocytes (FLS), immune cells, cytokines, chemokines, and their regulation
 - Standard therapies (adalimumab, sarilumab, sirukumab, methotrexate [MTX], baricitinib)
 - Clinical outcomes (DAS28-CRP score)



Platform Qualification

The reference virtual patient (VP) is representative of an average moderate/severe MTX-inadequate responder RA patient with stable, chronic disease

• Cell numbers in blood and synovium were calibrated to match the average from several published studies for moderate/severe RA patients (Figure 3) • Synovial mediator levels, determined by clearance rates, cell numbers, and cell-type specific production rates also matched reported literature averages



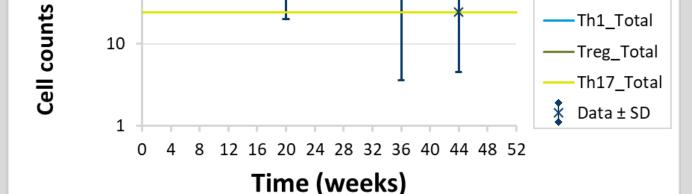


Figure 3. Blood cell concentrations (left) and synovial cell densities (right) in untreated reference VP. Baseline Platform calibration represents a dynamic equilibrium that reflects stable, chronic disease. Data values are plotted at arbitrary time points.

Serial histological studies from biopsies of patients were used to qualify cellular responses to therapies

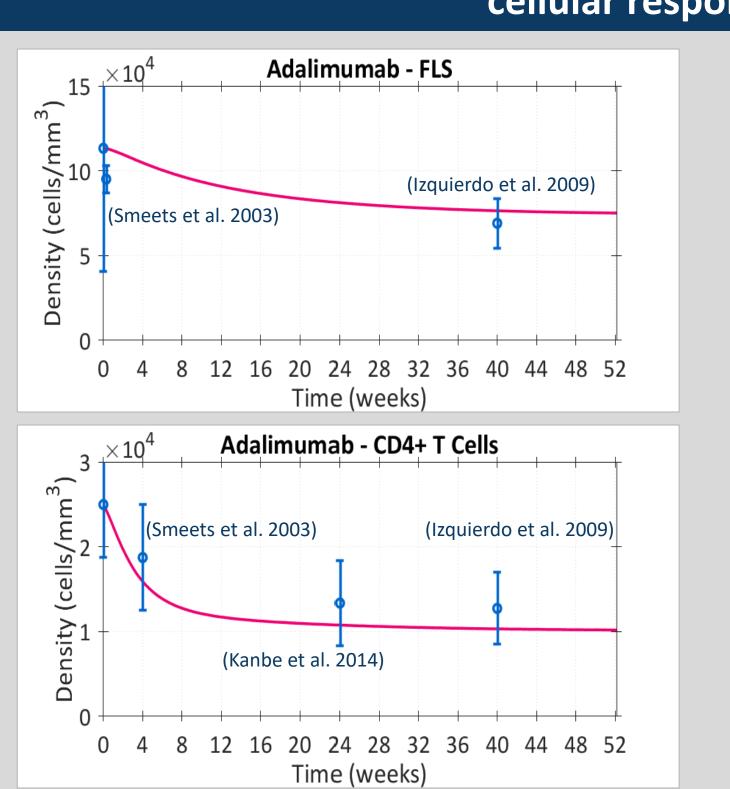


Figure 4. Reduction in synovial FLS, macrophages, CD4⁺ T cells and B cells in anti-TNFα treated reference VP. Simulations (pink line) of 40 mg Q2W adalimumab treatment in reference VP compared to published data (**\$\phi\$**: mean ± SD)



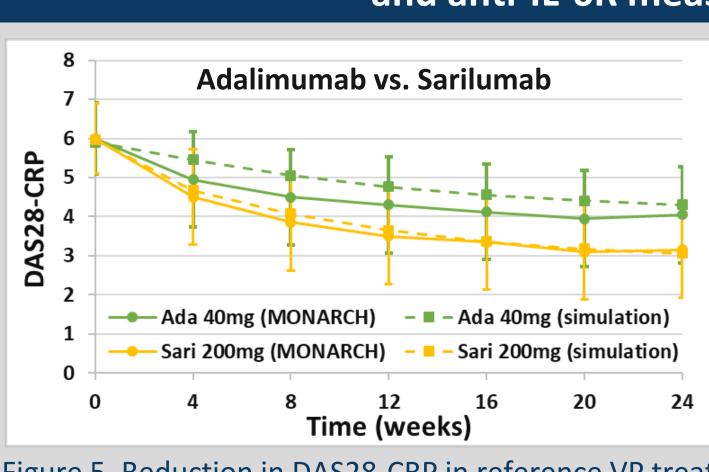


Figure 5. Reduction in DAS28-CRP in reference VP treated with anti-TNF α or anti-IL-6R (left) or anti-IL-6R + MTX (right). Simulations in reference VP (dashed lines) of 40 mg Q2W adalimumab, 200 mg sarilumab Q2W or 150 mg sarilumab Q2W + MTX vs. clinical score from the MONARCH (Burmester et al. 2017) and MOBILITY trials (Genovese et al. 2015) (**\phi**: mean ± SD)

Reference VP has an average clinical response to all implemented test therapies in combination with MTX

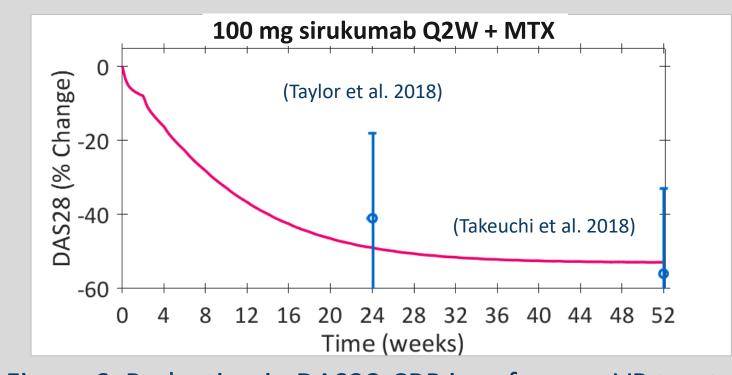
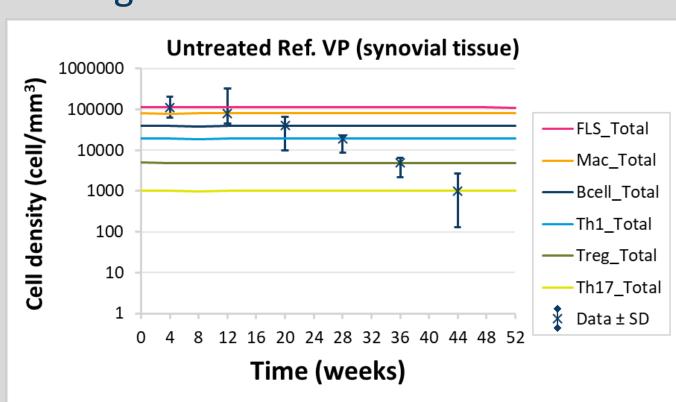
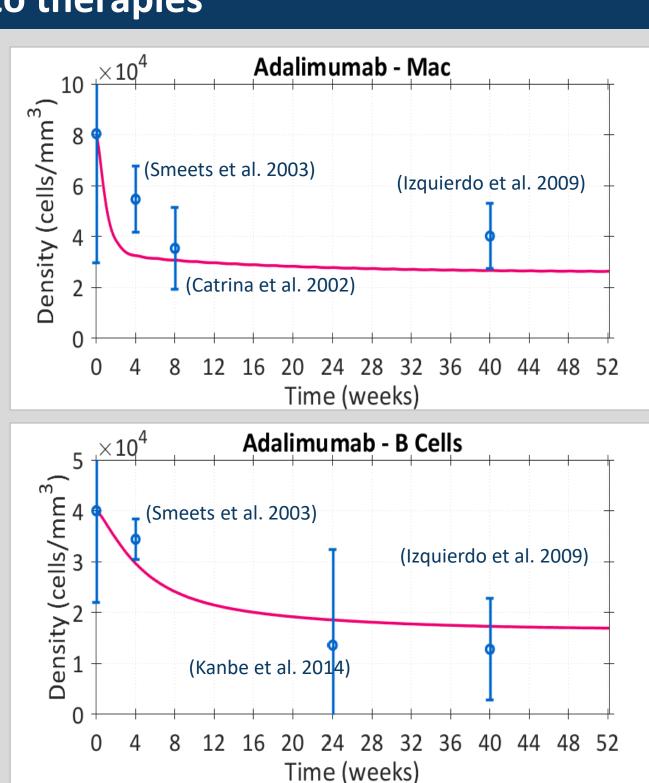
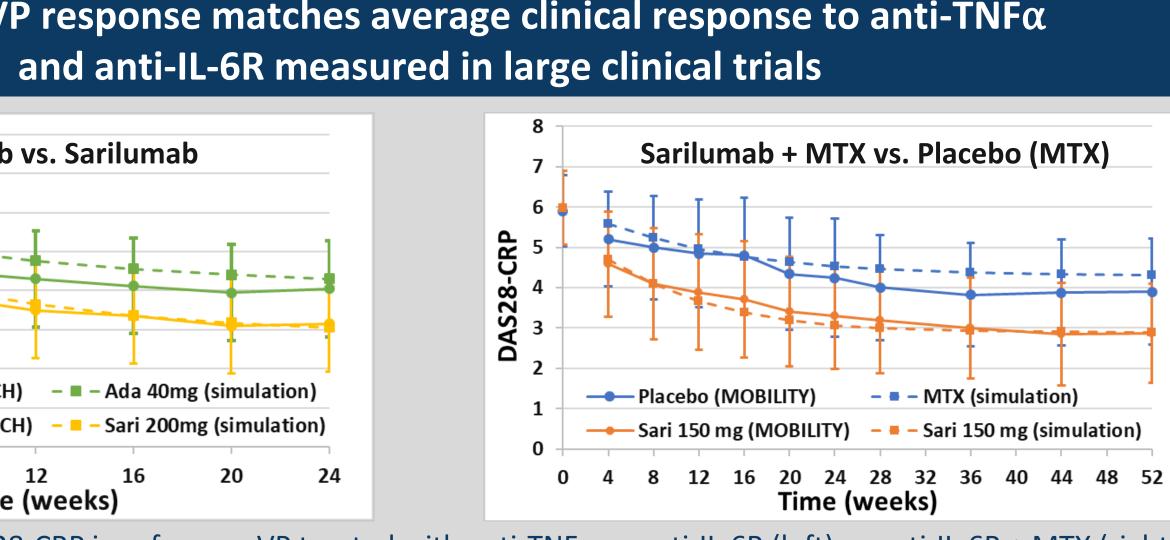


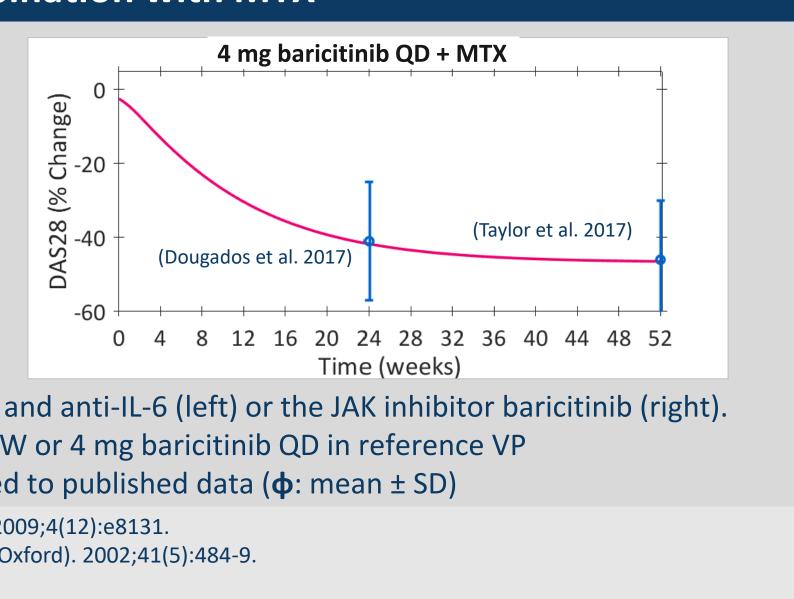
Figure 6. Reduction in DAS28-CRP in reference VP treated MTX and anti-IL-6 (left) or the JAK inhibitor baricitinib (right). Simulations (pink line) of 100 mg sirukumab Q2W or 4 mg baricitinib QD in reference VP

on a MTX-background treatment compared to published data (ϕ : mean ± SD) 4. Izquierdo E, Canete JD, Celis R, Santiago B, Usategui A, Sanmarti R, et al. PLoS One. 2009;4(12):e8131 5. Catrina AI, Lampa J, Ernestam S, af Klint E, Bratt J, Klareskog L, et al. Rheumatology (Oxford). 2002;41(5):484-9. Kanbe K, Chiba J, Nakamura A. Rheumatol Int. 2014;34(1):125-30. 7. Burmester GR, Lin Y, Patel R, van Adelsberg J, Mangan EK, Graham NM, et al. Ann Rheum Dis. 2017;76(5):840-7. 8. Genovese MC, Fleischmann R, Kivitz AJ, Rell-Bakalarska M, Martincova R, Fiore S, et al. Arthritis & rheumatology (Hoboken, NJ). 2015;67(6):1424-37. 9. Taylor PC, Schiff MH, Wang Q, Jiang Y, Zhuang Y, Kurrasch R, et al. Ann Rheum Dis. 2018. 10. Takeuchi T, Tanaka Y, Yamanaka H, Harigai M, Nakano T, Akagi K, et al. Modern rheumatology. 2018:1-9. 11. Dougados M, van der Heijde D, Chen YC, Greenwald M, Drescher E, Liu J, et al. Ann Rheum Dis. 2017;76(1):88-95. 12. Taylor PC, Keystone EC, van der Heijde D, Weinblatt ME, Del Carmen Morales L, Reyes Gonzaga J, et al. New England journal of medicine. 2017;376(7):652-62. 13. Dennis G, Jr., Holweg CT, Kummerfeld SK, Choy DF, Setiadi AF, Hackney JA, et al. Arthritis research & therapy. 2014;16(2):R90.









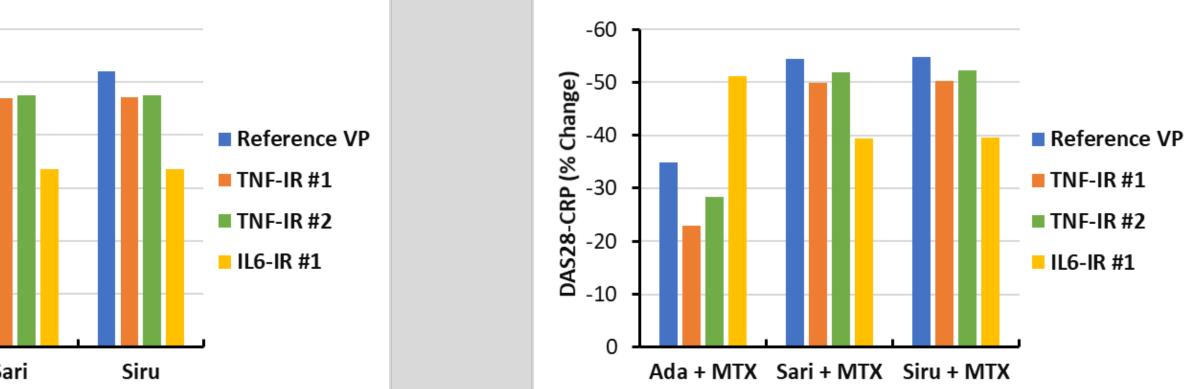
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	• •	racteristics of the fou	ir major phenotypes	s of RA synovium	identified
	Phenotype	Lymphoid	Myeloid	Low inflammation	Fibroid
	Defining	B and/or T	chemotaxis, TNFα and	inflammatory	TGFβ & bone
	gene clusters (microarrays)	lymphocyte activation and differentiation, Ig	IL-1β production, phagocytosis,	response and wound response	morphogenetic protein signaling,
		production, IL-17 signaling	mononuclear cells proliferation	processes	endocytosis
	Synovial cell	• T cells: ++	• T cells: +++	• T cells: ++	• T cells: ++
	infiltration (histology,	 B cells: +++ Mac.: +++ 	 B cells: + Mac.: +++ 	 B cells: - Mac.: ++ 	 B cells: - Mac.: ++
	FACS) Serum	Fibroblasts: +sICAM: low	Fibroblasts: ++sICAM: high	Fibroblasts: ++sICAM: low	 Fibroblasts: +++ sICAM: low
	biomarkers	• CXCL13: high	• CXCL13: med	• CXCL13: med	• CXCL13: low
Fo	our alternate VI		phenotypes w α and anti-IL-6		-
-	Table 2. Characteristi				
				Response to th	• •
	VP phenotype		MTX	Anti-TNFo	
-	Reference VP Anti-TNF-IR #1*		±	++ ±	+++
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*	IR : inadequate-respon	e VPs cover a ra			
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• IL-6 pathways are key disease drivers in a majority of the VPs phenotypes resulting in more patients responding to anti-IL-6R than to anti-TNF α therapies



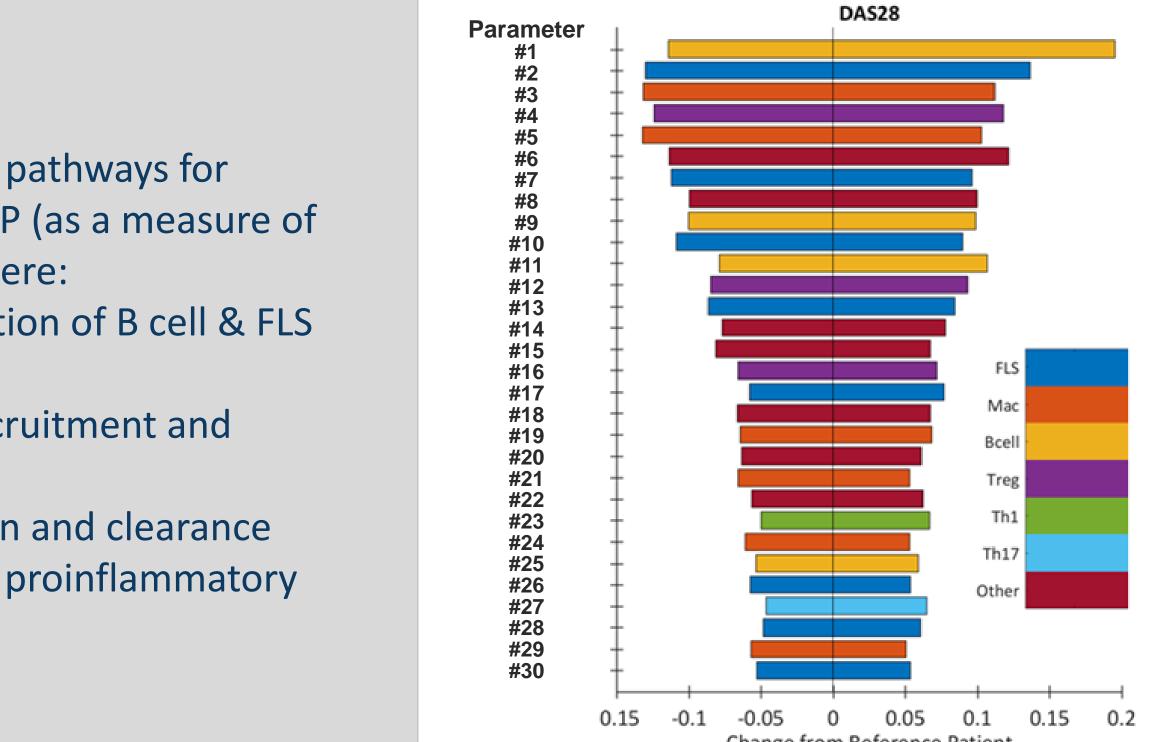
Alternate Virtual patients

and anti-



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or anti-

For more information about this work, please contact:

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